

Dale Bumpers College of Agricultural, Food & Life Sciences Animal Science





College of Veterinary Medicine Driven to Discover



Practices of -omics techniques in swine production improvement and probiotics development.

Presented by: Dr. Xiaofan Wang

October 22th 2021



Content

Introduction of multi-omics and their applications in gut microbiome study

Swine probiotics: market size and economic potential

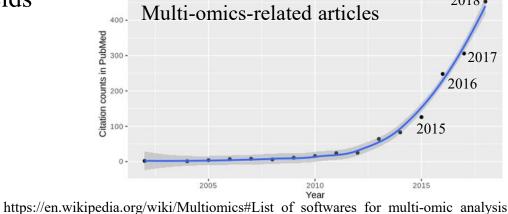
Characterization of longitudinal swine gut microbiome using culture dependent and independent approaches

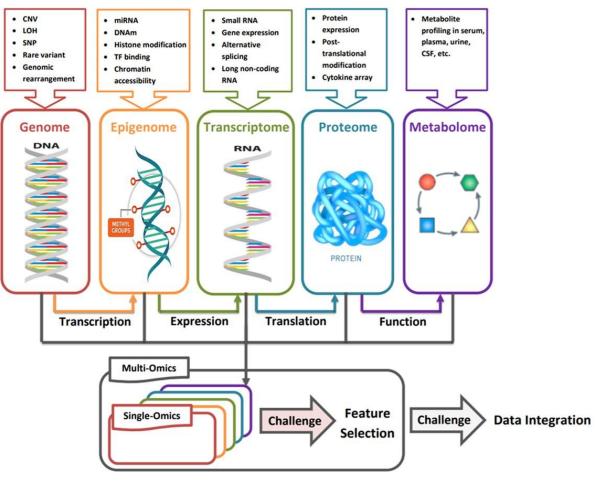
Probiotics identification, isolation, and validation using multi-omics methods



Multi-omics and their applications in gut microbiome study

- a high-throughput sequencing based biological analysis approach consisting of microbiomics (16s rRNA, ITS, culturomics, and metagenomics), genomics, transcriptomics, epigenomics, proteomics, metabolomics, and patholomics etc.
- quantitatively track changes in biological processes in unprecedented detail (Ali Ebrahim et al., 2016)
- disclose relationships between biological entities and identify biomarkers characterizing biological systems.
- rapid growth in size and complexity of multi-omics data sets indicated the advent of big-data era in biological and medical fields
 Multi omics related articles



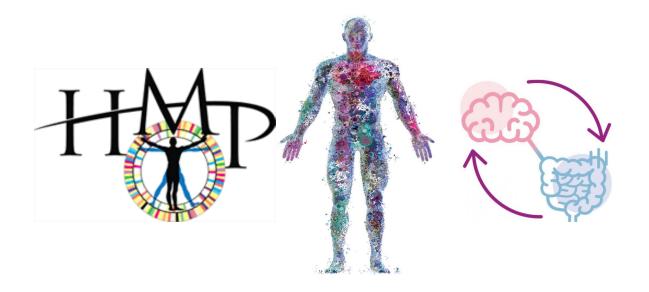


(Momeni et al., 2020)



Multi-omics and their applications in gut microbiome study

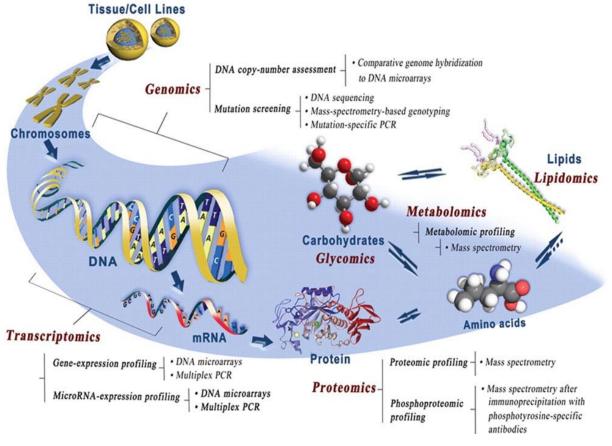
- Applications
- Human Microbiome Project
 - Phase 2: integration of multi-omic data from host/microbiome to human disease
 - Gut/nasal microbiome and type 2 diabetes
 - Gut microbiomes and inflammatory bowel disease
 - Vaginal microbiomes and pre-term birth
 - Brain-gut axis
- Systems Immunology
 - collect omics data that related to immune response to infectious diseases such as pediatric chikungunya
 - effectiveness and side effects of vaccines (transcriptome and plasma metabolomics)
- Cancer
 - multi-omics data repositories (Subramanian et al., 2020)
- Precision medicine



DATA REPOSITORY	WEB LINK	DISEASE	TYPES OF MULTI-OMICS DATA AVAILABLE
The Cancer Genome Atlas (TCGA)	https://cancergenome.nih.gov/	Cancer	RNA-Seq, DNA-Seq, miRNA-Seq, SNV, CNV, DNA methylation, and RPPA
Clinical Proteomic Tumor Analysis Consortium (CPTAC)	https://cptac-data-portal. georgetown.edu/cptacPublic/	Cancer	Proteomics data corresponding to TCGA cohorts
International Cancer Genomics Consortium (ICGC)	https://icgc.org/	Cancer	Whole genome sequencing, genomic variations data (somatic and germline mutation)
Cancer Cell Line Encyclopedia (CCLE)	https://portals.broadinstitute. org/ccle	Cancer cell line	Gene expression, copy number, and sequencing data; pharmacological profiles of 24 anticancer drugs
Molecular Taxonomy of Breast Cancer International Consortium (METABRIC)	http://molonc.bccrc.ca/ aparicio-lab/research/ metabric/	Breast cancer	Clinical traits, gene expression, SNP, and CNV
TARGET	https://ocg.cancer.gov/ programs/target	Pediatric cancers	Gene expression, miRNA expression, copy number, and sequencing data
Omics Discovery Index	https://www.omicsdi.org	Consolidated data sets from 11 repositories in a uniform framework	Genomics, transcriptomics, proteomics, and metabolomics

Multi-omics and their applications in gut microbiome study

Technologies and data analysis methods



https://www.pratiks.info/research/harvard/novel-multiomics-technologies-to-decipher-molecular-signatures-of-infectious-diseases and the second seco

A community-maintained list of software packages for multi-omics data analysis.

Website: <u>https://github.com/mikelove/awesome-multi-omics</u>

- Multi-omics correlation or factor analysis
- Ecology multi-table literature
- Chemometrics multi-table literature
- Behavioral research multi-table literature
- Multi-omics clustering / classification / prediction
- Multi-omics networks
- Single cell multi-omics
- Multi-study correlation or factor analysis
- Multi-omics reviews / evaluations
- Multi-omics application papers
- Multi-omics data management

Multi-omics networks

- 2018 MolTi-DREAM Didier identifying communities from multiplex networks, and annotated the obtained clusters article
- 2019 RWR-MH Valdeolivas Random walk with restart on multiplex and heterogeneous biological networks article

Animal probiotics and future market size

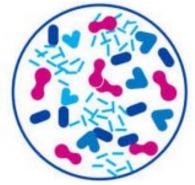




Probiotics Live microorganisms that confer a health benefit on the host when administered in adequate amounts.

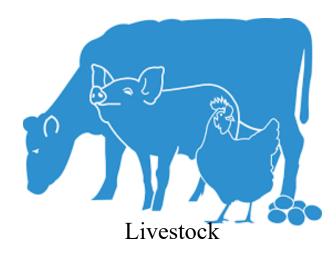
Prebiotics Substrates that are selectively utilized by both probiotics and host microorganisms, conferring a health benefit.

Synbiotics Combination of prebiotics



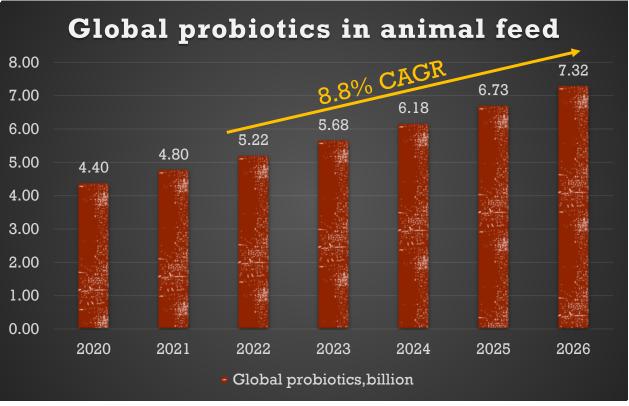
Postbiotics Compounds produced by microorganisms, released from food components or microbial constituents, including non-viable cells that when administered in adequate amounts, promote health and well-being.





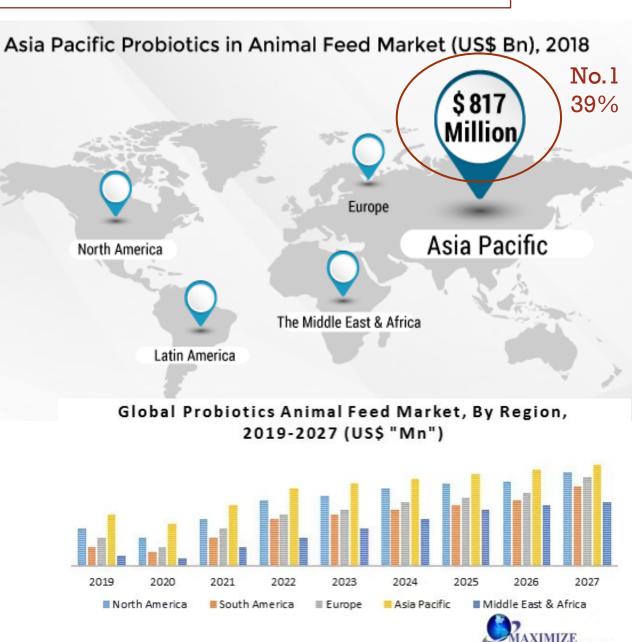


Animal probiotics and future market size



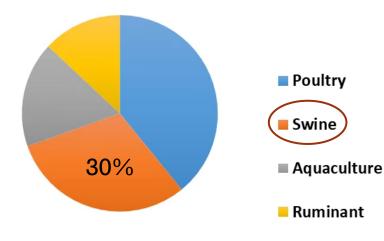
https://www.prnewswire.com/news-releases/probiotics-in-animal-feed-market-worth-7-3-billion-by-2026--exclusive-report-by-marketsandmarkets-301354278.html

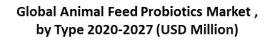
- The China Ministry of Agriculture and Rural Affairs released the ban of antibiotics use in animal feed from 2020.
- The Asia Pacific is projected to account for the largest market size during the forecast period.

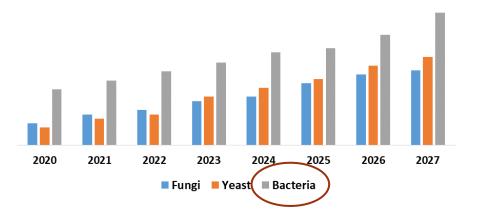


Swine probiotics

Global Animal Feed Probiotics Market, by Installation type (%) in 2020

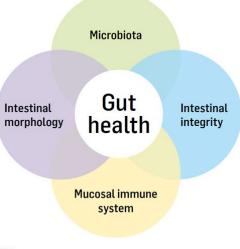






- Three traditional swine probiotics: *Lactobacillus*, *bacillus*, and yeast
 - ✤ Benefits:
 - improve gut homeostasis, alleviate stress, and preventing diarrhea
 - ✤ Immune protective and pathogen prevention
 - ✤ Improve feed efficiency and meat production
 - Drawbacks:
 - Inconsistent or poor growth promoting performance
 - Limited probiotics options (GRAS list)
 - Un- or inadequately characterized biological functions and delivery mechanism

A workflow combining culture-dependent and cultureindependent approaches using multi-omics techniques was suggested for revolutionary probiotic selection in swine industry.



BIOMIN



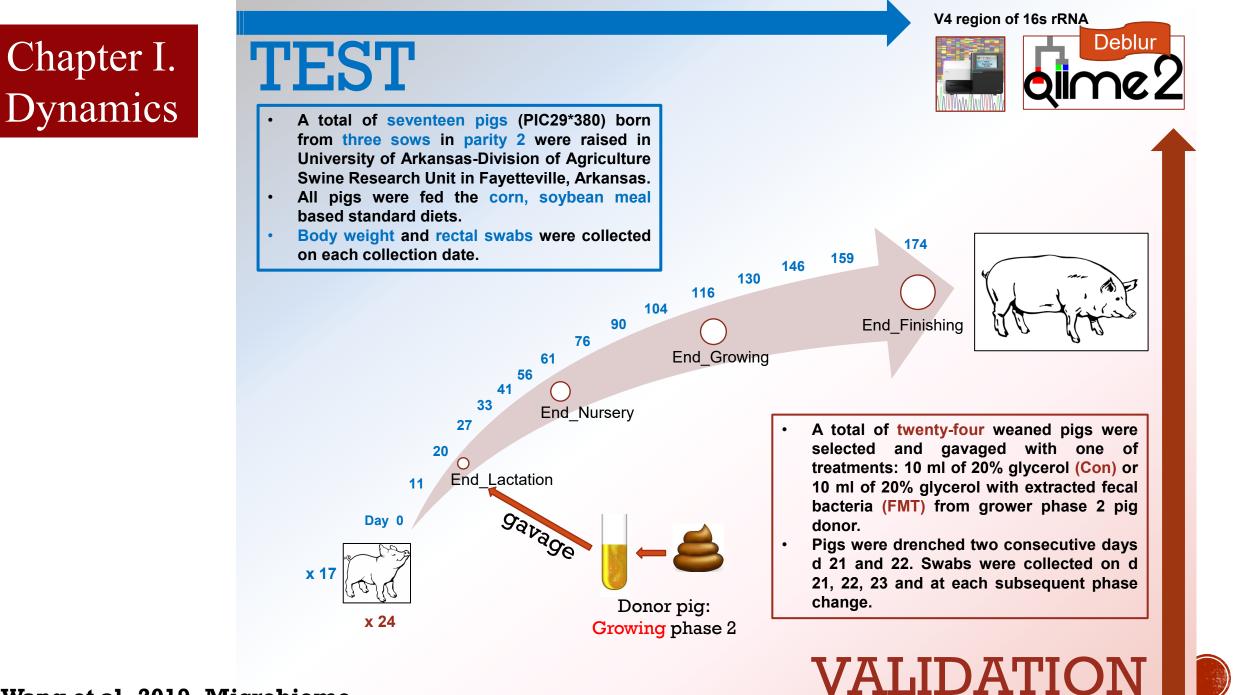
Characterization of longitudinal swine gut microbiome using culture dependent and independent approaches

KEY BIOLOGICAL AND ECOLOGICAL QUESTIONS

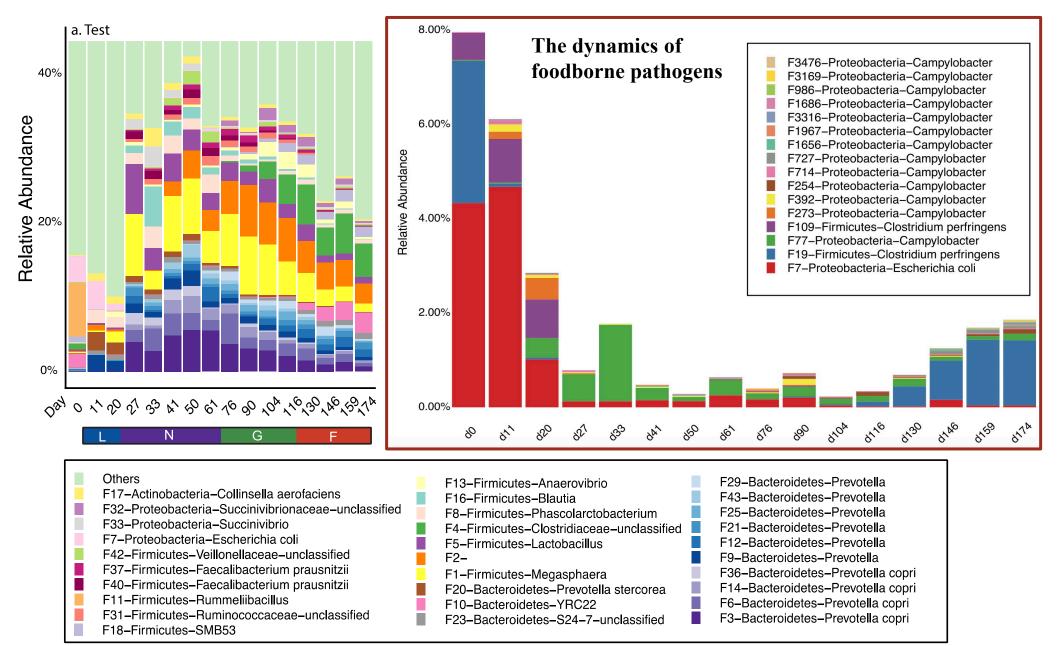
- How does the swine gut microbiome change from birth to market?
- Whether adult pigs' microbiota can colonize in young piglets?
- What are the major drivers shaping the swine gut microbiome?
- Who are the residents and who are passengers?
- Is there a core microbiome?
- Are probiotics stage-related?
- What is the dynamics of pathogens in swine gut?







Dynamics of the swine gut microbiome from birth to market: composition





Growth performance associated bacteria at different growth stages

		Test study		
Lactation % 25.9	Nursery % 69.21	Growing % 56.43	Finishing % 42.44	Overall % 93.14
Lactation % 25.9 $F77$ \circ \circ $F323$ \circ \circ $F363$ \circ \circ $F190$ \circ \circ $F190$ \circ \circ $F450$ \circ \circ $F450$ \circ \circ $F450$ \circ \circ $F264$ \circ \circ $F876$ \circ \circ $F360$ \circ \circ $F360$ \circ \circ $F295$ \circ \circ $F162$ \circ \circ $F295$ \circ \circ $F162$ \circ \circ $F295$ \circ \circ $F162$ \circ \circ $F270$ \circ \circ $F270$ \circ \circ $F270$ \circ \circ $F270$ \circ \circ $F249$ \circ \circ $F249$ \circ \circ $F340$ \circ \circ </td <td>F234 O F17 O F47 O F222 O F202 O F347 O F1 O F1 O F202 O F347 O F1 O F604 O F2 O F205 O F195 O F7 O F104 O F223 O F379 O F221 O F319 O F439 O F333 O F233 O F12 O F439 O F333 O F233 O F12 O F439 O F333 O F12 O F48 O F545 O F154 O F36 O F283</td> <td></td> <td>$\begin{array}{c ccccccccccccccccccccccccccccccccccc$</td> <td>$\begin{array}{c ccccccccccccccccccccccccccccccccccc$</td>	F234 O F17 O F47 O F222 O F202 O F347 O F1 O F1 O F202 O F347 O F1 O F604 O F2 O F205 O F195 O F7 O F104 O F223 O F379 O F221 O F319 O F439 O F333 O F233 O F12 O F439 O F333 O F233 O F12 O F439 O F333 O F12 O F48 O F545 O F154 O F36 O F283		$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$
F138 0 F714 0 F65 0 F141 0	F336 0 F274 0 F242 0 F301 0	F464 0 F446 0 F348 0 F75 0	F353 0 F102 0 F99 0 F192 0	F121 0 F309 0 F381 0 F202 0
F21 0 F123 0 F172 0 F172 0	F299 0 F35 0 F13 0	F196 0 F269 0 F74 0	F81 0 F342 0 F584 0	F231 0 F1 0 F376 0 F42 0





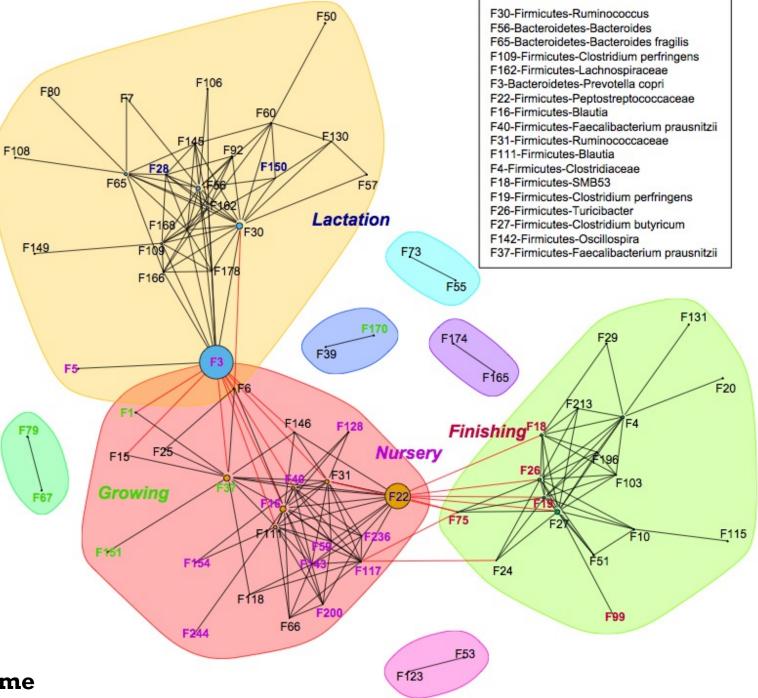


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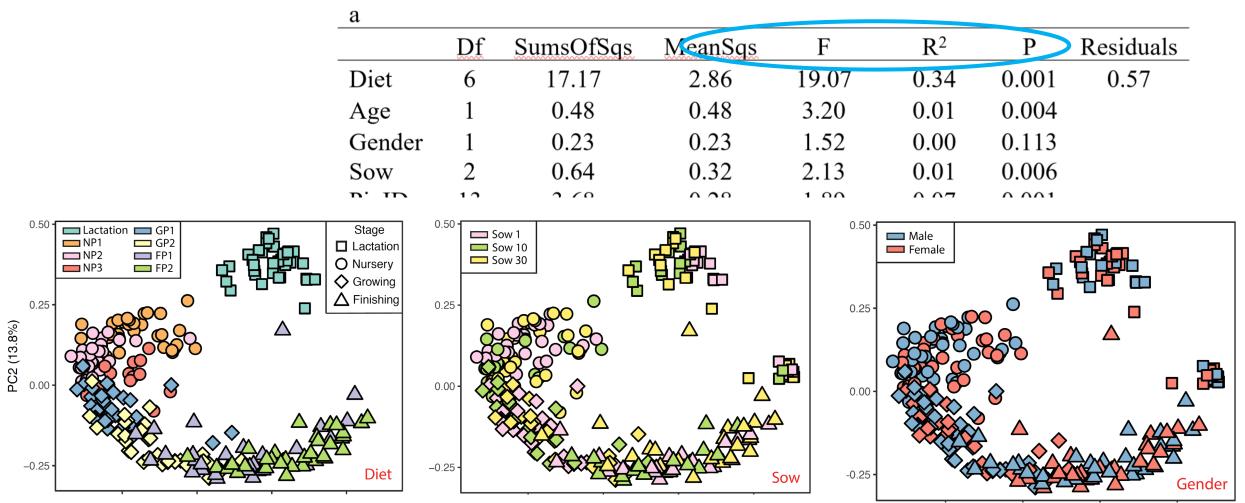
"Passenger"

Co-occurred bacteria/ networks



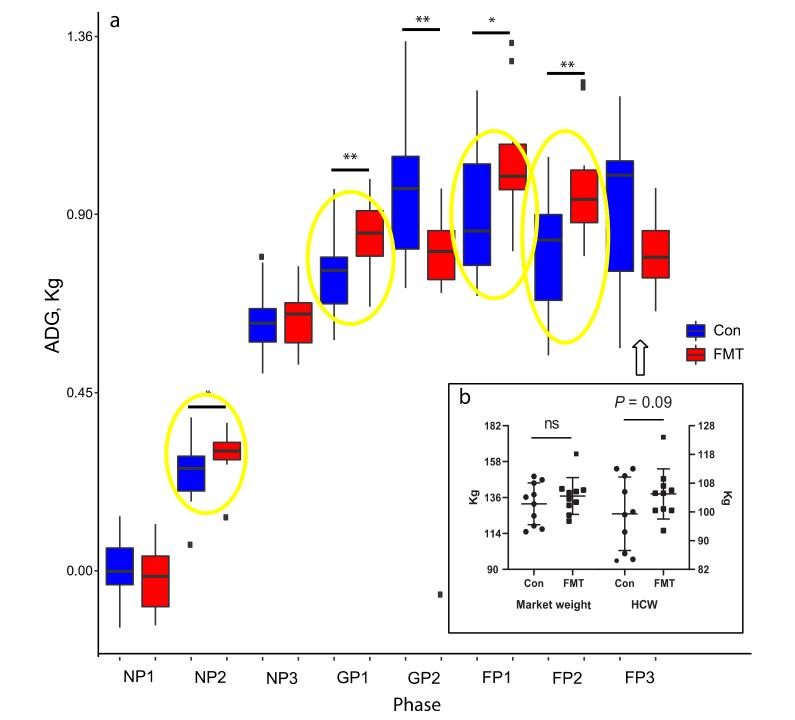


What are the drivers of gut microbiome?



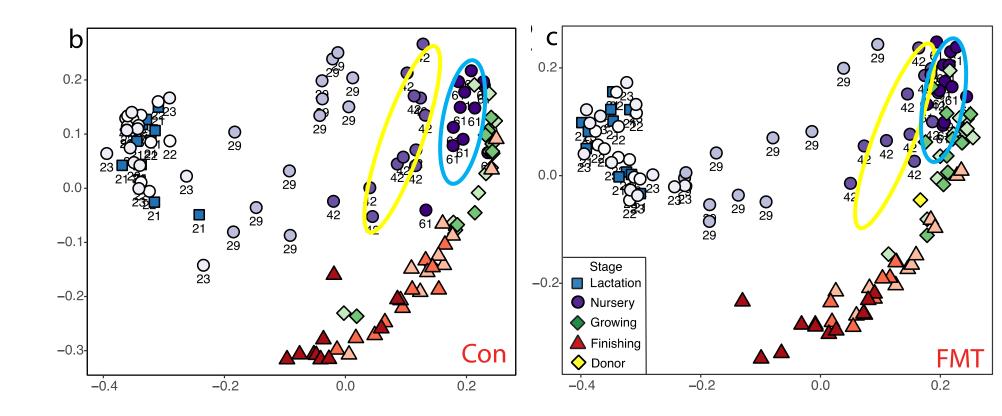


Growth performances



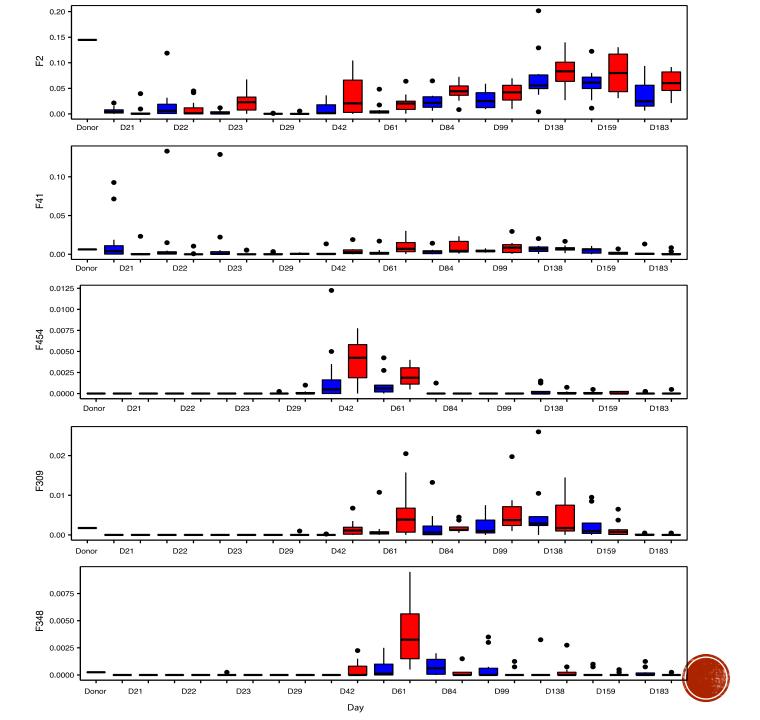
Jaccard

Different microbiome structures on D42 and D61



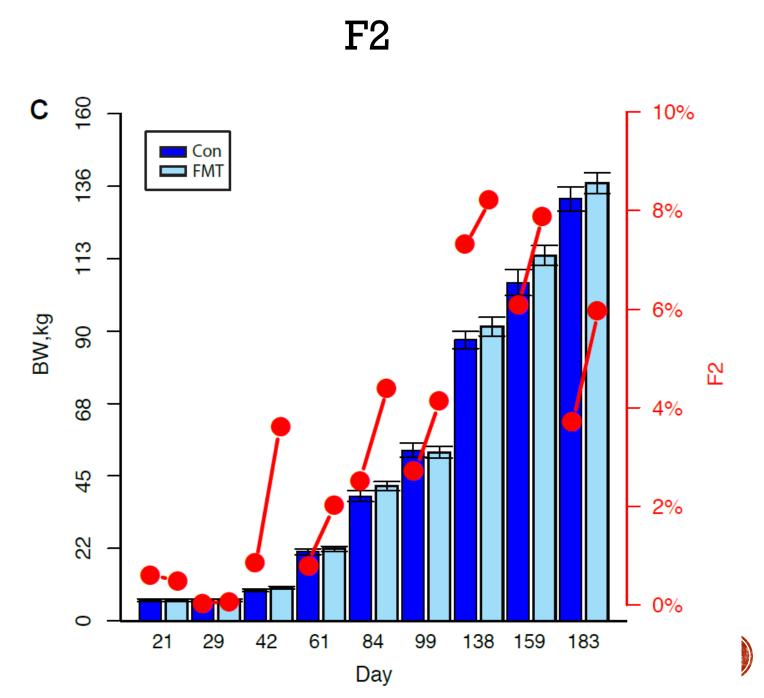


"Colonized" in FMT group



Potential probiotic





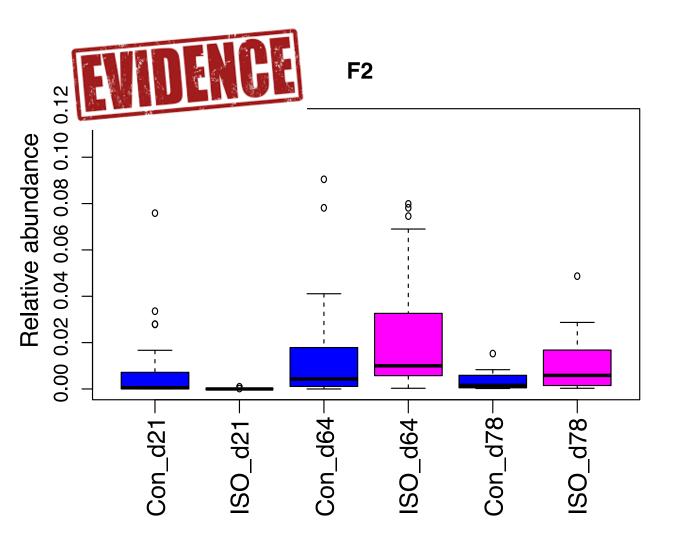
Growth performance associated bacteria at different growth stages

Potential probiotic

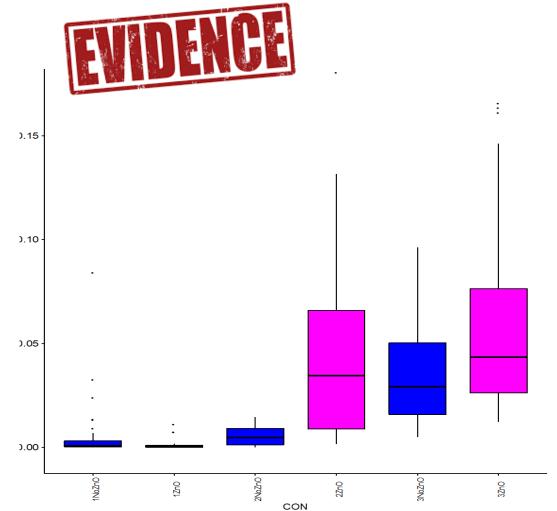


					Test study				
Lacta	ation % 25.9	Nurs	ery % 69.21	Grow	ing % 56.43	Finis	hing % 42.44	Overa	all % 93.14
F77 F323 F363 F190 F182 F450 F264 F876 F360		F234 F17 F47 F222 F202 F347 F1 F1 F2		F4 F18 F26 F27 F121 F309 F22 F127 F445		F75 F26 F295 F19 F261 F631 F22 F27 F309		F55 F4 F27 F75 F19 F26 F234 F18 F47	C C C C C C C C C C C C C C
F295 F162 F694 F35 F1 F270 F298		F205 F195 F7 F104 F223 F379 F221	· · O · · · · · · · · · · · · · · · · ·	F100 F382 F330 F13 F484 F473 F395	0 0 0 0 0	F377 F184 F128 F116 F215 F37 F333	0 0 0 0 0	F73 F64 F395 F336 F174 F184 F261	0 0 0 0 0
F168 F22 F2 F130 F253 F340	0 0 0 0 0	F319 F439 F333 F233 F12	· O · · · · · · · · · · · · · · · · · ·	F215 F73 F505 F188 F581	0 0 0 0	F114 F119 F329 F245 F231 F381		F165 F222 F127 F103 F377 F295	
F249 F413 F446 F346 F166	0 0 0 0	F48 F545 F154 F339 F36 F245		F40 F414 F103 F381 F231 F29		F100 F258 F61 F262 F194	0 0 0 0	F114 F196 F631 F223 F22	0
F257 F92 F317 F224 F502 F138		F42 F180 F283 F228 F348 F336	0 0 0 0 0	F338 F66 F433 F19 F335 F464	0 0 0 0 0 0	F297 F115 F117 F111 F371 F353	0	F37 F195 F433 F545 F379 F121	
F714 F65 F141 F21 F123 F172		F274 F242 F301 F299 F35 F13		F446 F348 F75 F196 F269 F74		F102 F99 F192 F81 F342 F584	0	F309 F381 F202 F231 F1 F376 E42	0

Relative abundances of F2 in the isolator trial



Relative abundances of F2 in pigs fed with or without ZnO

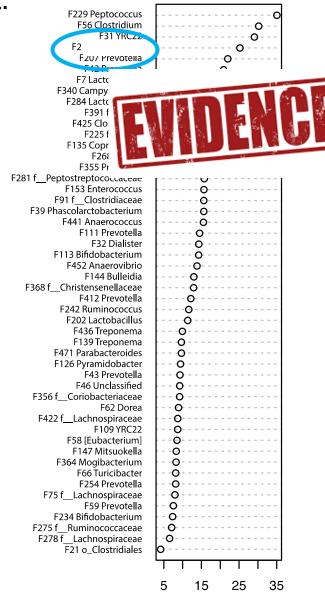


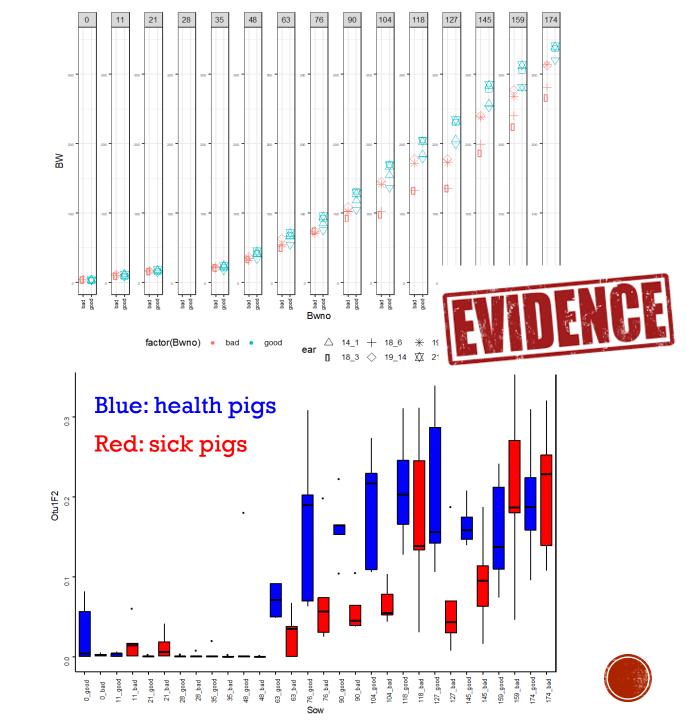
Tsai et al., 2018, Frontiers in Microbiology

Wei et al., 2020, microorganisms



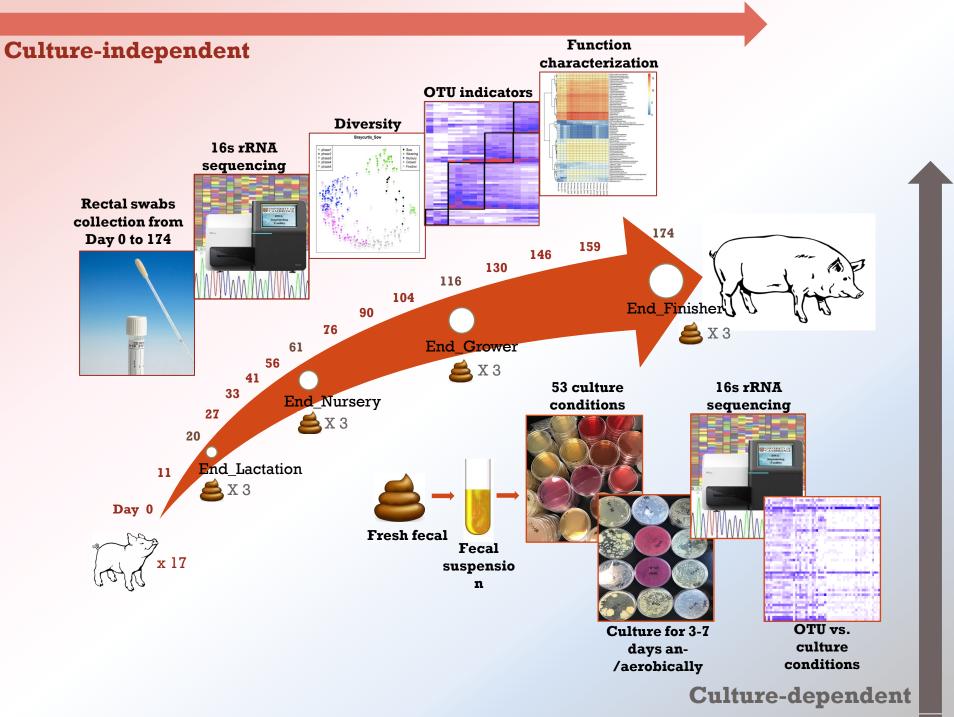
The top 50 estimated age (EMA)-associated features isolated by the regression based random forest.



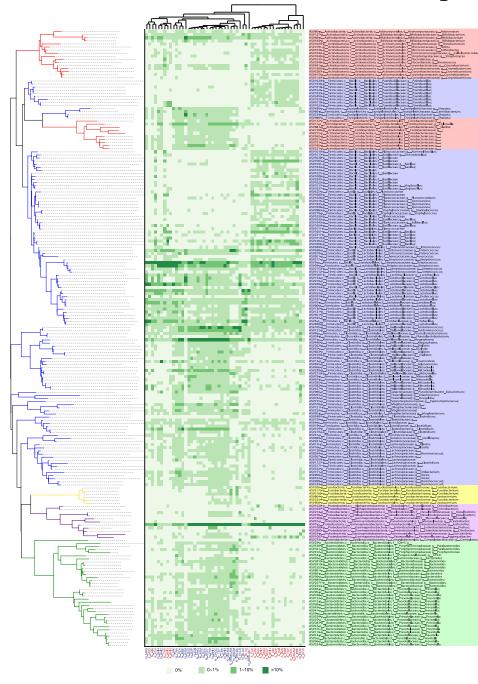


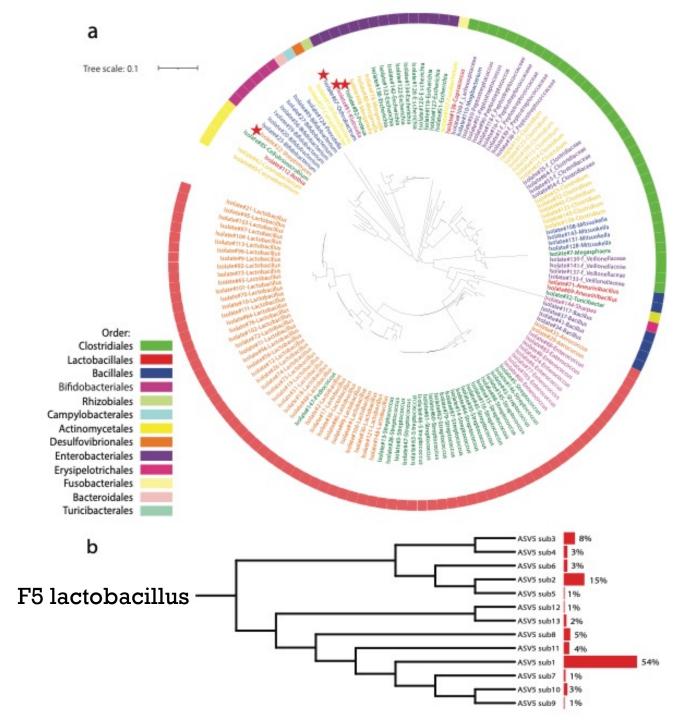
Culturomics experimental design

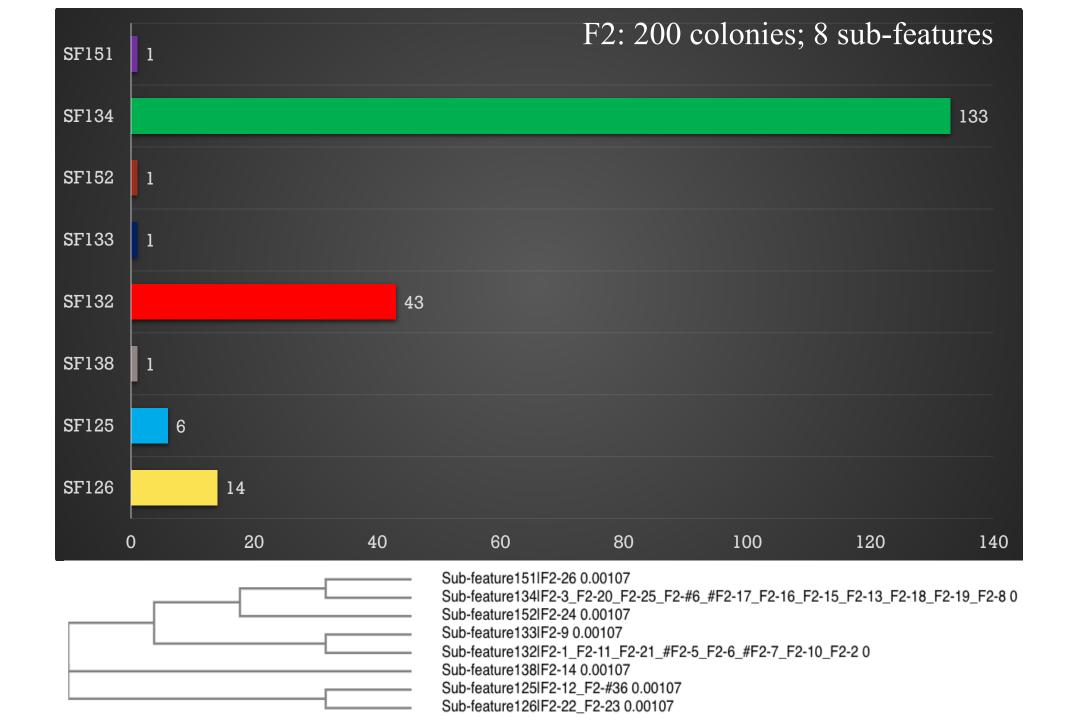
Culturomics



ASV-culture method heatmap

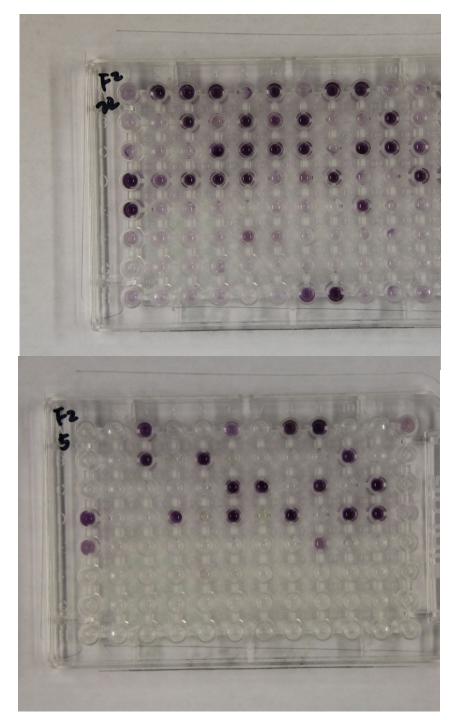


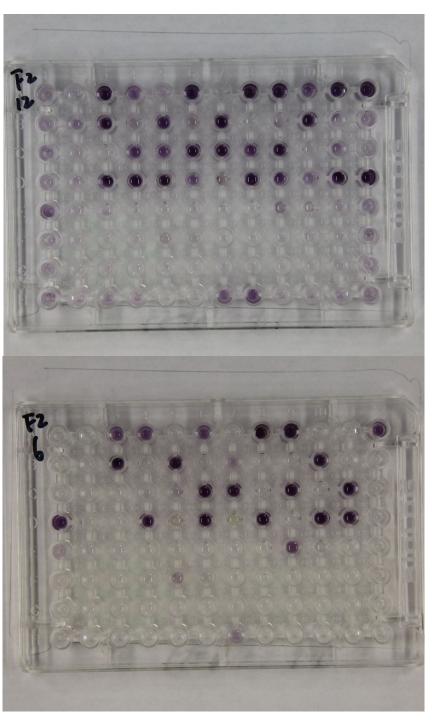


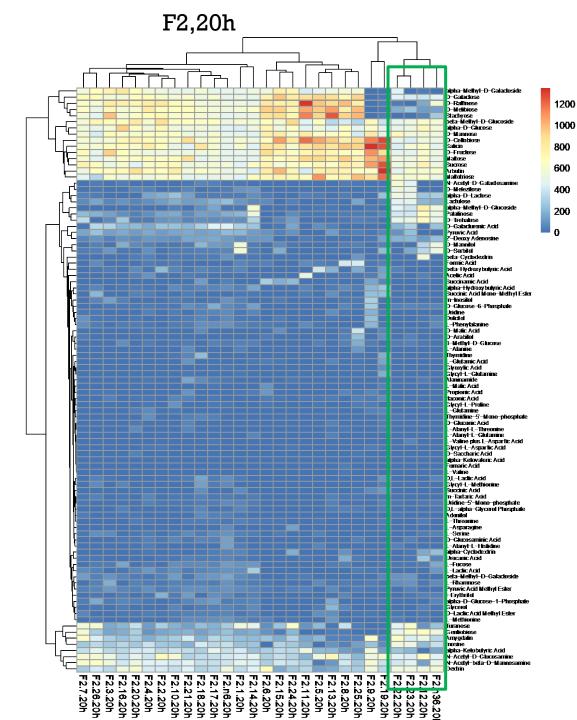


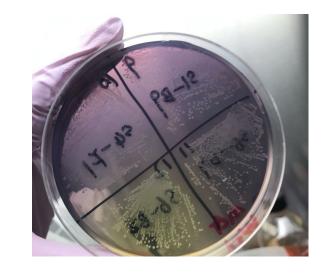


Metabolic profile

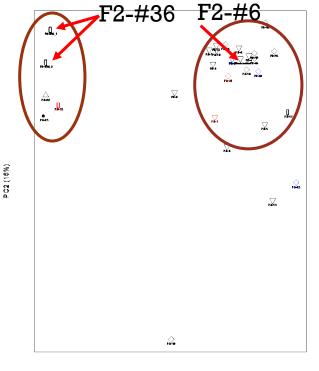






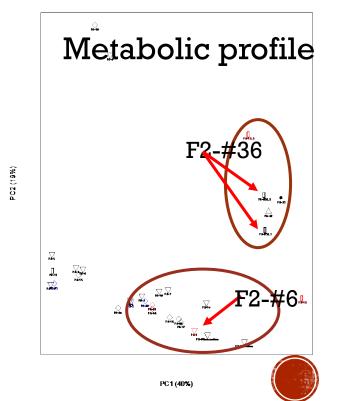


Bray-curfis_4h

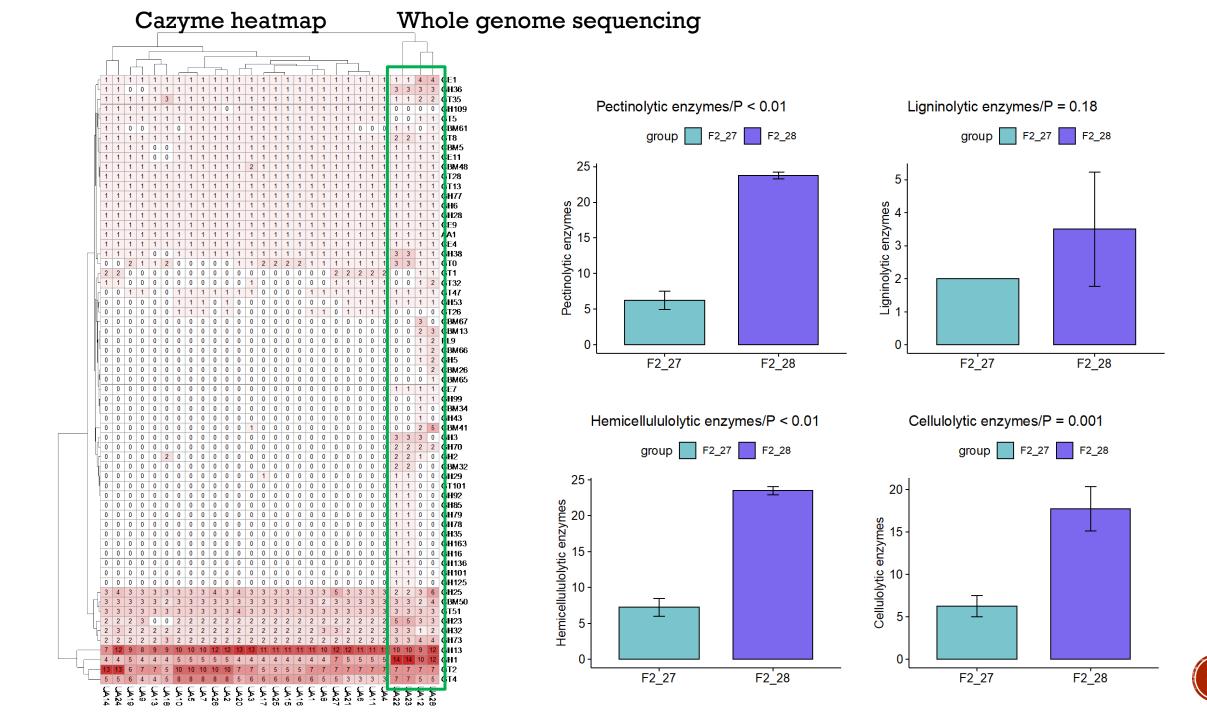




Bray-curtis_20h



PC1 (40%)



F2 oral inoculation trial 1:

Body weight of pigs from all six treatments

_		lb kg lb kg <th< th=""></th<>																									
	Age	2	.5	34	4	42		53		6	6	8	1	9	4	10	38	12	22	13	36	1	50	16	54	17	78
		lb	kg	lb	kg	lb	kg	lb	kg	lb	kg	lb	kg	lb	kg	lb	kg	lb	kg	lb	kg	lb	kg	lb	kg	lb	kg
	2.F2_Str1	0.3	0.2	1.2	0.5	3 1	1.4	1.7 (0.8	4.2	1.9	4.1	1.8	5.2	2.4	12	5.4	14	6.4	17	7.5	20	9.1	21	9.3	25	12
	3.F2_2	0	0	0.7	0.3		1.4	1.9 0	0.9	4.4	2	7.7	3.4	-1	-0	6	2.7	7.5	3.4	7	3.1	4.4	2	6.8	3.1	14	6.3
	4.F2_Str3																										
	5.Lact_1								_																		
l	6.Lact_2	0.2	0.1	1.5	0.7	3 1	1.3	1.6 0	0.7	3	1.4	5.3	2.4	0.3	0.2	6.5	2.9	9.5	4.3	19	8.4	19	8.6	21	9.5	21	9.7
	20	2	5	34	1	42		53		66	6	8	1	9	4	10)8	12	22	13	36	15	50	16	54	17	8
300 200 Mg 100			Str1 Str2 Str3 t_1												÷								I i ↓ i				*
(2 - N										φ.= N									5-5			
	1.Con 2.F2_Str1 3.F2_Str2 4.F2_Str2 5.Lact_1 6.Lact_2	1.Con 2.F2_Str1 3.F2_Str2	4.F2_Str3 5.Lact_1 6.Lact_2	1.Con 2.F2_Str1 3.F2_Str2	4.F2_Str3 5.Lact_1 6.Lact_2	1.Con 2.F2_Str1 3.F2_Str2 4.F2_Str3	5.Lact_1 6.Lact_2	1.Con 2.F2_Str1 3.F2_Str2 4.F2_Str3	5.Lact 1 6.Lact 2	1.Con 2.F2_Str1 3.F2_Str2	4.F2_Str3 5.Lact_1 6.Lact_2	1.Con 2.F2_Str1 3.F2_Str3	4.F2_Str3 5.Lact_1 6.Lact_2	1.Con 2.F2_Str1	5.Lact_2 6.Lact_2	1.Con 2.F2_Str1 2.E2_Str1	5.Lact_2 6.Lact_2 6.Lact_2	1.Con 2.F2_Sh1	4. 5.Lact 1 6.Lact 2								

F2 oral inoculation trial 2: F2-#28

D) and D1: F2 (gavage		D21: F2 gava	ge			• Metag	RNA gene genome colom	e sequen	cing	
			Nursery						g	Finishing		
	DO	D7	D14	D21	D28	D40	D77	D104	D126	D134	D162	
	11/11/20	11/18/20	11/25/20	12/2/20	12/9/20	12/21/20	1/27/21	2/23/21	3/17/21	3/25/21	4/22/21	
Trt 1 Con	13.7	13.73	14.8		26.18	43.34	42.63	64.11	92.95	107.33	134.97	
Trt 2 F2	13.57	13.78	15.12		29.03	46.08	44.74	69.60	99.79	110.62	141.34	
Trt 3 Bacillus	13.73	13.75	14.4		25.66	41.14						
Trt 4 F2+B	13.48	13.68	14.79		28.45	46.35						
Trt 5 B2	13.64	13.59	14.61		26.8	43.48						
Trt 6 Antibiotics	13.68	14.19	14.86		28.82	45.5	45.24	71.36	99.68	113.82	140.74	
	Î	Î	Î			Î	Î				ass performa	
				_						2. Tissu •	e collection Pancreas ti	

BW and rectal swabs

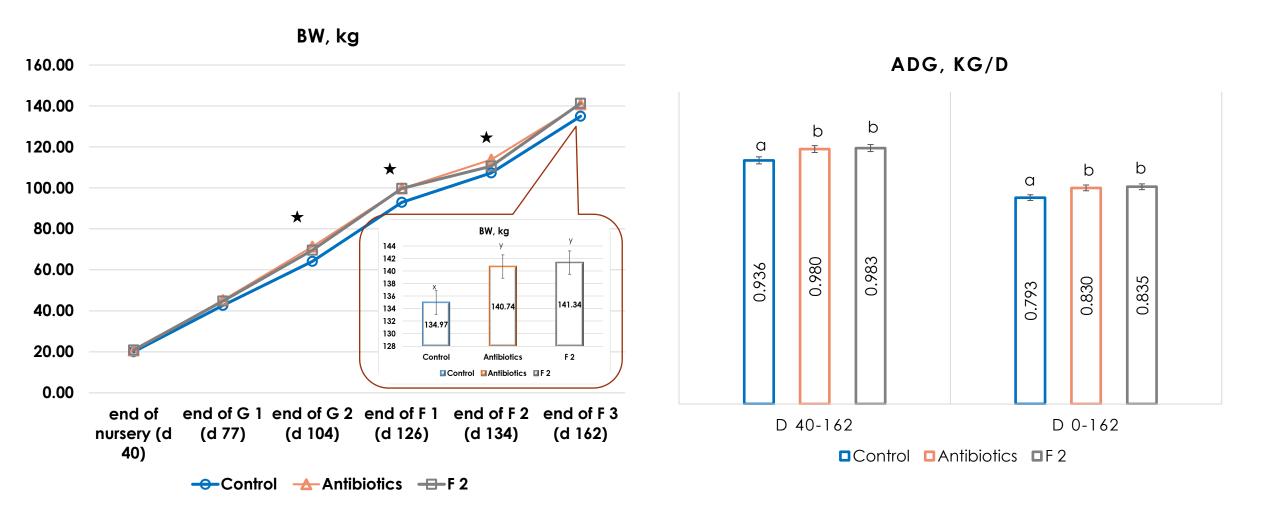
16S rRNA

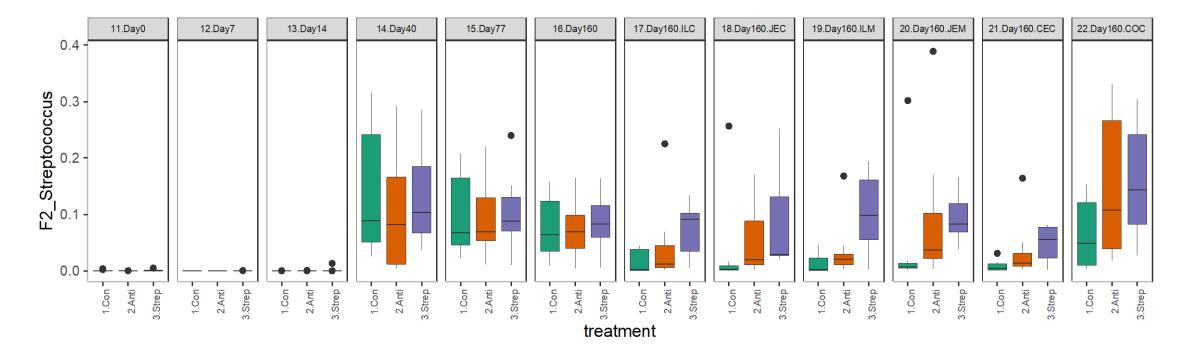


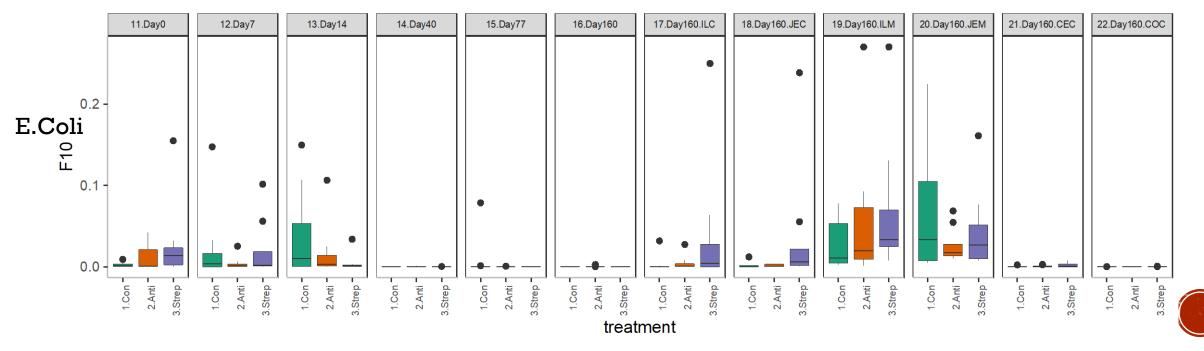
Enzyme activity •

Ileum and jejunum tissue, mucosa, and digesta content Cecum and colon tissue,

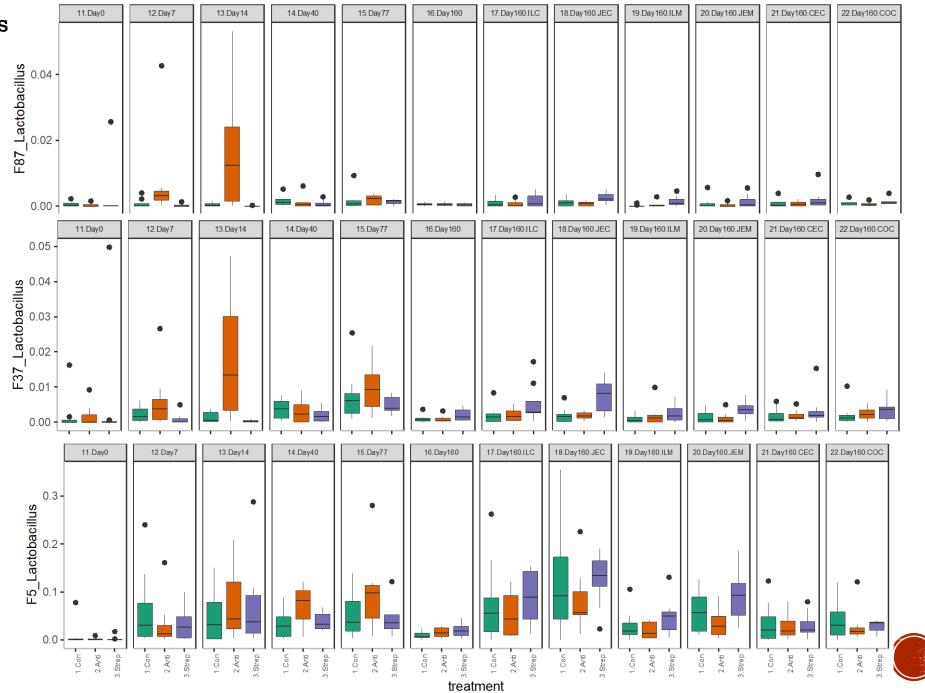
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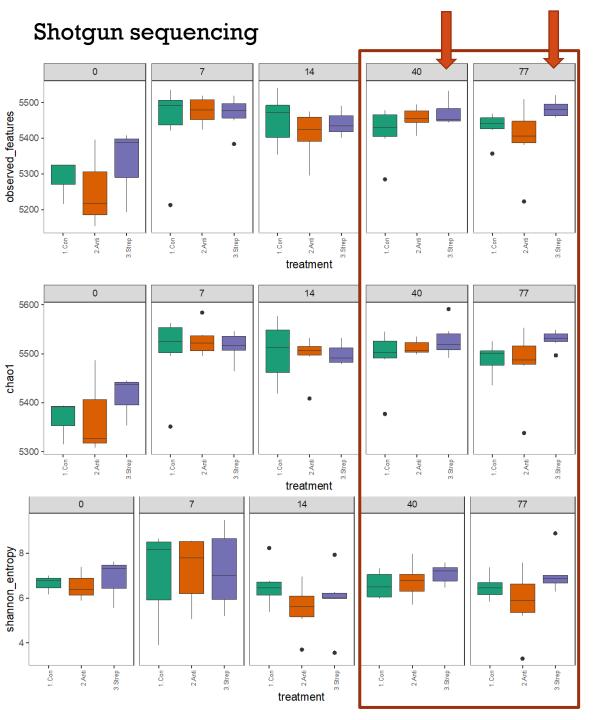


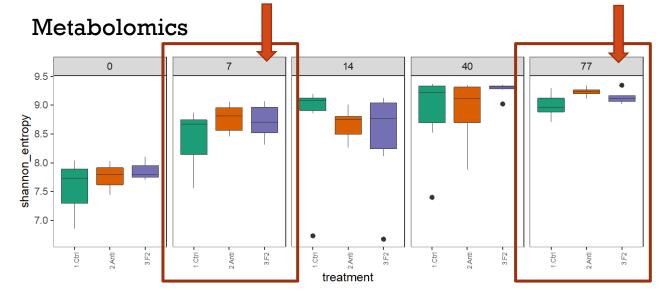


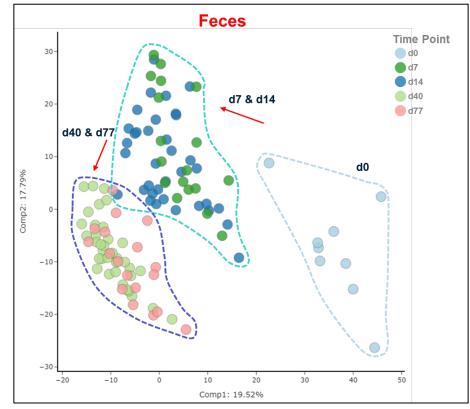


Modulation of lactobacillus in the gut











Metabolomics-d14

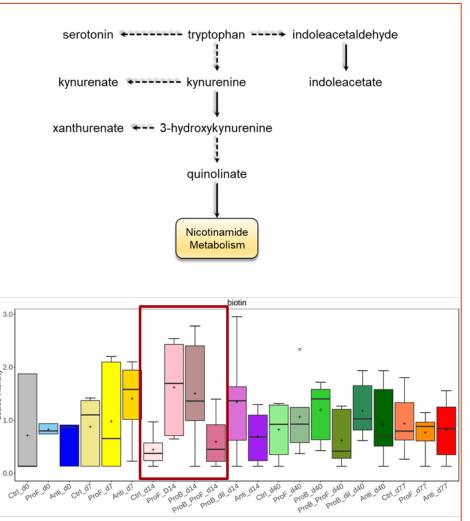
					Benzoate Metabolism
	[Fold of Change		Aromatic Organic Acids Amino Acids
		Welch's Two	Two Way Repe	atad Maaauraa	Organic Acids CAmino Acids
Sub	Biochemical Name	Sample t-Test	ANC		Polyphenols Dietary Aromatics
Pathway		PROB D14	PROBIOTICF D14	ANTIBIOTIC D14	
		CTRL D14	CONTROL D14	CONTROL D14	
	phenylalanine	0.90	0.96	1.07	
Phenylalanin	N-acetylphenylalanine	1.23	0.83	2.22	cholesterol
e Metabolism	N-methylphenylalanine	1.90	1.96	1.14	benzoate benzoate
	2-hydroxyphenylacetate	1.28	1.58	1.22	Primary Bile Acids
	tyrosine	0.86	0.92	1.03	Bile Acids
T	tyramine	0.70	0.18	0.45	catechol opio hippurate
Tyrosine Metabolism	tyrosol	1.25	0.52	0.86	
	vanillactate	0.39	0.59	0.82	catechol of hippurate "Energy" hippurate Primary Bile Acids Primary Bile Acids Bile Acid
	gentisate	0.30	0.40	0.70	"Energy"
	tryptophan	1.04	0.93	0.92	Secondary Bile Acids
	N-acetyltryptophan	1.25	1.21	1.31	catechol
	oxindolylalanine	1.59	1.37	1.32	sulfate
Tryptophan	kynurenine	1.14	1.18	1.41	
Metabolism	kynurenate	0.54	0.50	0.78	Digestive Lumen
	anthranilate	1.43	1.32	1.37	
	tryptophol	0.50	0.60	1.20	
	indolepropionate	0.61	0.69	0.71	Excretion
	skatol	1.54	1.12	0.36	
	6-beta-hydroxylithocholate	1.82	1.68	0.50	
	lithocholate	1.37	1.39	0.47	indolepropionate
	tauroursodeoxycholate	0.74	0.08	0.49	
	dehydrolithocholate	1.57	0.89	0.80	3.0
Secondary	hyocholate	0.56	1.53	0.90	
Bile Acid	glycohyocholate	0.40	0.16	0.27	
Metabolism	taurohyocholate*	0.93	0.11	1.20	
	hyodeoxycholate	0.76	0.95 0.09	0.59 0.16	≥2.0
	taurohyodeoxycholic acid	0.13	0.09	0.16	
	glycohyodeoxycholate glycocholenate sulfate*	0.12	0.18	0.05	
	7alpha-hydroxycholestenone	1.73	1.79	1.26	
					$\overset{\circ}{}_{1.0}$



Metabolomics-d14

0.0. Ctrl_d0

			Fold of Change			
Sub Pathway	Biochemical Name	Welch's Two Sample t-Test	Two Way Repe ANC			
-		PROB D14	PROBIOTICF D14	ANTIBIOTIC D1		
		CTRL D14	CONTROL D14	CONTROL D14		
	nicotinate	0.70	0.71	0.98		
Nicotinate and	nicotinamide	1.20	0.82	0.64		
Nicotinamide	1-methylnic otinamide	0.29	0.17	0.42		
Metabolism	2-hydrox ynicotinate	0.43	0.54	0.38		
	trigonelline (N'-methylnic otinate	0.02	0.02	0.04		
Pantothenate and CoA	pantoate	2.73	1.14	2.80		
Metabolism	pantothenate	0.86	0.73	1.31		
Ascorbate and Aldarate	threonate	0.64	0.39	1.16		
Metabolism	oxalate (ethanedioate)	0.70	0.83	1.02		
	alpha-toc opherol	1.25	1.39	1.07		
	alpha-tocopherol acetate	1.65	1.81	1.33		
Tocopherol Metabolism	delta-toc opherol	1.47	1.59	1.26		
	alpha-tocotrienol	1.17	1.47	1.01		
	gamma-toc otrienol	1.26	1.40	1.01		
Biotin Metabolism	biotin	3.40	3.66	1.55		
Thiamine Metabolism	thiamin (Vitamin B1)	1.79	1.34	0.91		
Vitamin A Metabolism	retinol (Vitamin A)	3.01	2.07	2.14		
	pyridoxine (Vitamin B6)	1.32	0.95	0.84		
Vitamin B6 Metabolism	pyridox amine	1.37	1.37 1.17			
	pyridox ate	1.19	0.33	1.64		
	nicotin	ate				







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